

## Dialysis of Antibodies

Please note – this protocol has been optimised for certain conditions and is meant as a guide only, you may need to alter some parameters to suit your own experiment

1. Wearing gloves, cut a piece of dialysis tubing, 20 cm is sufficient for up to 5 ml of antibody.
2. Place the tubing in a glass beaker containing about 500 ml of distilled water.
3. Boil for 10 minutes (eg in microwave on high for 15 mins)  
Alternatively the whole roll of dialysis tubing can be boiled in this fashion and then stored at 4°C in sterile PBS/Azide for use as required.
4. Allow to cool a little, then remove tubing from water (wearing gloves) and tie a knot in one end. Pull the knot tight to prevent leakage.
5. Rub the open end of the tubing between fingers to open it up, then use a plastic pipette (glass may tear the tubing) to transfer the antibody solution into the tubing, making sure it runs down inside the tubing and doesn't bubble over the top
6. Ensure as much of the solution is sitting at the bottom of the tubing by running fingers downwards along the tubing, then expel as much air as possible by running fingers upwards along the tubing. Tie off the open end as close to the solution as possible.
7. Place sealed tubing in a beaker containing 1 - 2 litres of appropriate solution and place on stirrer in cold room
8. Dialyse for the rest of the day, then replace solution in beaker and dialyse overnight
9. Following dialysis, cut one end off the tubing and remove the solution using a plastic pipette.

**Note:** This procedure is adequate to get antibodies from one solution to another. However, dialysis of other proteins (ie significantly smaller or larger than antibodies) may require more extensive preparation of the dialysis tubing.